

Phytochemical Determination and Antimicrobial Investigation of some Selected Indian Medicinal Plants

Prathiba H.D¹ and N.H. Manjunath^{*2}

^{1,2}Department of Biochemistry, Central College Campus, Bangalore University, Bangalore-560001, (Karnataka), India
E-mail: ¹prateeba8@gmail.com, ²manjunath@gmail.com

Abstract: Use of plant based drugs and chemicals for curing various ailments and personal adornment is as old as human civilization. Recently, there has been growing interest in the traditional cures of livestock diseases because of the expensiveness of pharmaceutical products. The self-help approaches of traditional medicines, especially from medicinal plants offer a way by making use of the study was to find out the bioactive chemical constituents and to evaluate the antimicrobial activity of the methanolic extract of traditionally used five medicinal plants of India. A qualitative phytochemical analysis was performed for the detection of alkaloids, flavonoids, saponin and tannins, each analysis was carried out in triplicate which shows the positive result for alkaloids, flavonoids, saponin and tannins respectively, the highest yield of methanolic extract was found in *Daturastramonium* and *Lantana camara* in the alkaloid fraction. The antibacterial effect of some selected Indian medicinal plants was evaluated on bacterial strains like *Staphylococcus* species, *Escherichia coli* species and *Bacillus* species. The invitro antimicrobial activity was performed by disc diffusion method. The methanolic extract of *Daturastramonium* and *Lantana camara* showed the maximum activity against *Staphylococcus* species in alkaloids fraction compare to the flavonoid and saponin. The *Daturastramonium* showed the moderate zone of inhibition against the *Escherichia coli* species and *Bacillus* species. The flavonoid extract of the *Lantana camara* and *daturastramonium* showed the moderate zone of inhibition and the saponin showed the minimum zone of inhibition. However this work highlights its antibacterial sensitivity in par modern standard antibiotics. The result is quite encouraging, further studies needed.

Keywords: Medicinal plants, Phytochemicals, alkaloid, flavonoid, saponin, tannin, human pathogens, antimicrobial activity.

1. INTRODUCTION

In developing countries microorganism are frequently causes of prevailing diseases, presenting a serious public health issue in a significant segment of the population as uncovered by either private or official health care system (A.Rekha et al. 2011). Plants have formed the basis of sophisticated traditional medicine systems that have been in existence for thousands of years and continue to provide mankind with new remedies [Gloria E.Barboza et al., 2009]. In the recent years there has been an increasing awareness about the importance of medicinal plants [Dewick, P.M. et al., 1996]. According to WHO medicinal plants would be the best source to obtain

variety of drugs. Phytochemical which possess many ecological and physiological roles are widely distributed as plant constituents [Michael Wink et al., 2010]. The number of multi-drug resistant microbial strains and the appearance of strains with reduced susceptibility to antibiotics are continuously increasing. This increase has been attributed to indiscriminate use of broad-spectrum antibiotics, immunosuppressive agents, intravenous catheters, organ transplantation and ongoing epidemics of human immunodeficiency virus (HIV) infections (Dean DA and Gonzalez 1996). This situation provided the impetus to the search for new antimicrobial substances from various sources like medicinal plants (Cordell GA. et al., 2000). Synthetic drugs are not only expensive and inadequate for the treatment of diseases but are also often with adulterations and side effects. Therefore, there is a need to search for new infection-fighting strategies to control microbial infections (Sieradzki K, et al., 1999). Plant extracts have been used for centuries as a popular method for treating several health disorders. Numerous studies have been carried out on various natural products screening their antimicrobial activity. In the present work, qualitative phytochemical analysis was carried out in five plants [*Abutilonindicum*, *Adathodavisca*, *Daturastramonium*, *Lantana camara* and *tridax procumbent*] for the antimicrobial activity which have been known to possess medicinal property.

2. MATERIALS AND METHODS

2.1 Plant Collection and Identification:-

The plant materials *Adathodavisca*, *Daturastramonium*, *Lantana camara*, *tridaxprocumbens*, and *Abutilonindicum* were collected from fields in and around Bangalore city.

2.2 Preparation of Plant Material:-

The leaves plucked from the plant were washed 2-3 times with running tap water and was then air dried under shade. After complete shade drying the leaf material was ground in the mixer to obtain the powder and stored in plastic bags.

2.3 Extraction of Plant Material

Preparation of aqueous extracts:

Powdered leaf was homogenized with 5 gm of plant material was in 25 ml of water and the suspension was heat to 50-60 °c, and maintain for 15 minutes and then filtered. The filtrate was then centrifuged at 2500 rpm for 15 minutes and the clear supernatant was stored at 5° C until use.

2.4 Phytochemical Analysis

The phytochemical analyses were carried out according to the standard methods with minor modification. Phytochemicals analysis of the crude powder of the *Adathodaviscia*, *Daturastramonium*, *Lantana camara*, *tridaxprocumbens*, and *Abutilon indicum*, for the tests of phytochemicals as a alkaloid, saponin, tannins, flavonoides and protein etc were made as shown below.

2.4.1 Test for Alkaloides

200 mg plant materials were taken and added 10 ml Methanol and then filtered. After that 2 ml filtrate were taken and added 1 % HCL with steam 1 ml filtrate and 6 drops Mayer's reagent/Wagnersreagent/ Dragendorffsreagent. It produced creamish/Brown /Orange precipitate indicate the presence of alkaloids.

2.4.2 Test for Saponins

Approximate 0.5 ml filtered were taken and added 5 ml distilled water. Frothing persistence indicate presence of Saponin [GA Ayoola, HAB Coker].

2.4.3 Test for Tannins

200 mg plant material were taken and added 10 ml distilled water and then filtered. After that 2 ml filtered were taken and added 2 ml FeCl₃ Blue. Then black precipitate indicate the presence of Tannins & Phenols.

2.4.4 Test for Flavonoides

200 mg plant material were taken and added 10 ml Ethanol, then Tomato, Red colour indicate the presence of Flavonoids.

3. SELECTION OF BACTERIAL STRAINS

The investigated microorganisms consisted of Two Gram-positive bacteria: *Staphylococcus aureus* ATCC25923, *Bacillus subtilis* TCC6633; one Gram-negative bacteria: *Escherichia coli* ATCC25922. Microorganisms were obtained from the National Chemical Laboratory (NCL), Pune, India. Microorganisms were maintained at 4 °C on nutrient agar slants.

3.1 Antimicrobial susceptibility test

The antimicrobial assay was performed by agar disc diffusion method (Bauer AW, et al, . 1966). The 20ml of sterilized Muller Hinton Agar was poured into sterile petriplates, after solidification, 100 µl of fresh bacterial culture were swabbed on the respective plates. Each of discs which are approximately 5mm in diameter was cut from Whatman filter paper. The sterile discs were kept over the agar plates using sterile forceps at various concentration (2, 4, 6, 8, and 10µl). The plates were incubated for 24 hours at 37 °c. After incubation the diameter of inhibitory zones formed around each discs were measured (mm) recorded. (Nair R, KalariyaT, et al, .2005).

Table 1: Phytochemical Analysis of Crude Extract from the Leaves of Five Medicinal Plants

SL. NO	Phytoconstituents	Adathodaviscia	Abutilon indicum	Daturastramonium,	Lantana camara	Tridaxprocumbens
1	ALKALOIDS (a)Mayer's test (b)Dragendorff's test (c)Wagner's test	++ +	++ +	++ +	++ +	++ +
2	FLANOID S (a)Shinoda test (b)Alkline reagent (c)FeCl ₃ test	++ +	++ +	++ +	++ +	++ +
3	TANNINS (a)Lead acetate test (b)FeCl ₃ test	++	++	++	++	++
4	SAPONINS (a)Frathing test	+	+	+	+	+
5	AMINOACID (a)Millons test (b)Ninhydrin test	--	--	--	--	--
6	PROTEIN (a)Biuret test (b)Millons test	--	--	--	--	--

+ [Present] , - [Absent]

4. RESULT

The phytochemical analysis of the crude extract of the leaves of five medicinal plants revealed the presence of only alkaloids, flavonoids, tannin and saponin of all the phytochemical screened for anthraquinones, amino acid and protein were found to be absent (Table 1). The result obtained for the antimicrobial test performed on different extract of medicinal plants and zone of inhibition of the individual plant extract with three type of bacterial pathogen were identified that is *Escherichia coli*, *Bacillus subtilis*, *Staphylococcus aureus*.

Alkaloids antibacterial susceptibility test of five medicinal plants against some species of bacteria.

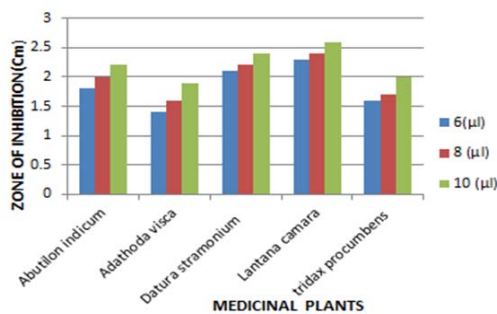


Fig. 1: *Escherichia coli*

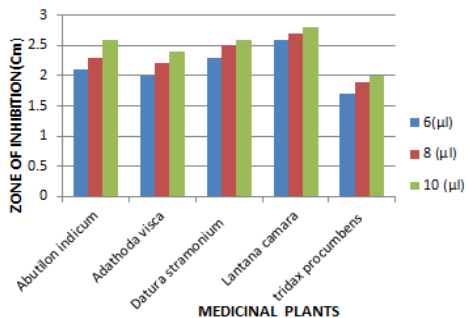


Fig. 2: *Staphylococcus aureus*

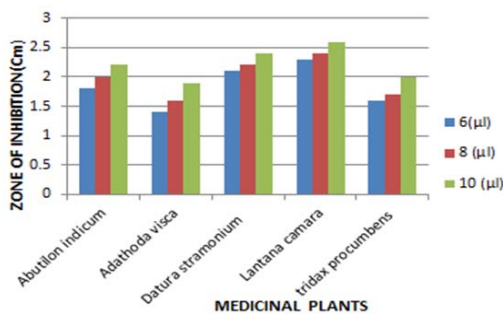


Fig. 3: *Bacillus subtilis*

Fig. 1: Represents the alkaloids antibacterial susceptibility test results for *Escherichia coli*, *Daturastramonium* and *Lantana camara* show the significance zone of inhibition that is the highest zone inhibition 2.0 cm was noted in the concentration of 10 µl. *Adathoda visca* and *tridaxprocumbens* 1.6 cm was noted in 10 µl. **Fig. 2:** Represents the alkaloids antibacterial susceptibility test results for *Staphylococcus aureus*, *Lantanacamara* show the significance zone of inhibition 2.8 cm was noted in the concentration of 10 µl and 2.7 cm noted in the 8 µl and 2.6 for 6 µl. *Daturastramonium* is the second highest zone of inhibition 2.6 cm was noted in the 10 µl.

Fig. 3: Represents the alkaloids antibacterial susceptibility test results for *Bacillus subtilis*. For *Lantana camara*, the highest zone of inhibition 2.6 cm was noted in the concentration of 10 µl, *Datura stramonium*, 2.4 cm was noted in the concentration of 10 µl, *adathodavisca*, 1.9 cm concentration of 10 µl, *Tridaxprocumbens* 1.8 cm concentration of 10 µl. From the above observation *Lantana camara* showed the significance zone of inhibition against the *Bacillus subtilis*. 1.1 cm in 8 µl and 1.2 cm was noted in the concentration of 10 µl. From the above observation *Lantana camara* showed the significance zone of inhibition against the *Escherichia coli*.

5. DISCUSSION

The increasing trend of the resistance to the antibiotics in current use has drawn the attention of researcher to natural alternative treatment of bacterial infection as potential source of new novel antibacterial agent. This study indicated that all the five plant extract were showing the antibacterial activity in different inhibition. The gram negative bacteria are reported to be resistant against most antibacterial agents as a result of the more complicated nature of their cell wall compared to gram positive bacteria (Akinyemi et al., 2005; Goyal et al., 2008). However the five species were found to be active against the two groups of bacteria underlining their ethno medicinal use for treatment of various infection diseases. The present study is an attempt to evaluate plants as source of potential chemotherapeutic agents and antimicrobial activity (Cledson, V. et al., 2007). The search of novel bioactive compounds including antimicrobial ones continues. This is largely so because some pathogens have developed resistance to certain currently used drugs and some disease have yet to be treated chemotherapeutically (Chin, Y. et al., 2006). Medicinal plants are necessary for the scientific point of view, to establish a rational relationship between chemical biological and therapeutical activities of folklore medicine (Harborne J.B, 1973). Our study stated that the antimicrobial activities of five medicinal plants (*Abutilon indicum*, *Adathodavisca*, *Daturastramonium*, *Lantana camara*, *tridaxprocumbens*) due to the presence of the following phytochemicals namely, alkaloids, flavonoids, Saponins and Tannins. The potential of antimicrobial properties of plants are related to their ability to synthesize compounds by the

secondary metabolism (Clark AM, et al, .1993). It has also been shown that tannins are biologically active, against *E. coli*, *S. aureus* and *Bacillus subtilis*. Results obtained from this study, indicated that, the plant extracts showed the strongest antimicrobial activity than the commercially available antibiotics. The antimicrobial activity of methanolic extract of *Daturastarmonium* and *lantana camara* showed the significant antimicrobial activity various phytopathogens.

6. CONCLUSION

The preliminary phytochemical screening and antimicrobial activity of five medicinal plants studied here can be seen as the potential source of useful drugs. Different extract showed varying degrees of antibacterial activities against microbes tested here. It also justifies the folklore medicinal uses and claims about the therapeutic values of this plant as curative agent and we therefore, suggest further, the purification and characterization of the phytochemicals that would be obtained with a view to obtaining useful chemotherapeutic agent. The plant studied here can be a source of high pharmacological importance and potential source of new drugs. Further studies on such bioactive compounds screening and their antimicrobial activity will unravel the potentiality of these traditional medicines.

REFERENCES

- [1] A.Rekha., B.Bharathi., et al, Effect of some medicinal herbal extracts on clinically important bacterial pathogens.IJPSR 2011;2(9):2362-2367.
- [2] Akinyemi K.O., Oladapo O, Okware C.E., et al, Screening of crude extract of six medicinal plants used in south west Nigerian unorthodox medicine for anti-methicillin resistant staphylococcus aureus activity. BMC Complement and Altern. Med., 5;6 (2005).
- [3] Bauer A. W., Kirby W.M.M., Sherris J.C., Antibiotic susceptibility testing by a standardized single disk method, Am J Clin Pathol. 45: 493-496, (1966).
- [4] Cordell G.A., Biodiversity and drug discovery - a symbiotic relationship, Phytochemistry 55: 463-480, (2000).
- [5] Cledson V., Simone M., ElzaSamnia F.A. and ArturSmania J.R., Screening methods to determine antimicrobial activity of natural products, Brazilian Journal of Microbiology. 38, 369-380, (2007).
- [6] Chin Y., Balaumas M.J., Chai H.B. and Kinghorn A.D., Drug discovery from natural sources, Journal of Ethnopharmacology , 8, 239-53, (2006).
- [7] Clark A.M. and Hufford C.D., Discovery and development of novel property antibiotics for opportunity infection related to the acquired immunodeficiency syndrome, (1993).
- [8] Dewick P.M.[1996] Tumor inhibition from plant; Tease and Evans.
- [9] GloriaE.Barboza et al., [2009] Medicinal plants: A general review and a phytochemical ethnopharmacological screening of the native argentine flora.v, 34.
- [10] GA Ayoola, HAB Coker], phytochemical screening and antioxidant activities of some selected Medicinal plants used for malaria therapy in southwestern Nigeria. j of pharmaceutical research, 2008;7 (3):1019-1024].
- [11] Goyal P, Khanna.A, Chauhan G, Kaushik .P. In vitro evaluation of crude extract of *Catharanthus roseus* for potential antibacterial activity. Int.J green pharm., 2;176-181.(2008).